

Cancer Nanomedicine: Advanced Nanocarrier Systems, Targeted Drug Delivery, Theranostics, and Emerging Strategies for Precision Oncology

Fouzan Arif Mulla¹, Mo Saad Sanaullah Khan¹, Irfan Nizamuddin Mansuri¹, Shirgar Abdul Basit Mo. Salman¹, Mahefuzabibi Mohammed Desai^{1*}

¹Shree Dhanvantary Pharmacy College near the railway station, Kim, Surat, Gujarat, India

***Corresponding Author E-mail: arafmahefuzaa@gmail.com**

Abstract:

Cancer remains one of the leading causes of morbidity and mortality worldwide despite substantial advances in diagnosis and treatment. Conventional therapeutic approaches, including chemotherapy, radiotherapy, surgery, and immunotherapy, are often limited by poor tumor selectivity, systemic toxicity, multidrug resistance, and inadequate drug accumulation at the disease site. Nanomedicine has emerged as a transformative strategy in oncology, offering innovative solutions for targeted drug delivery, improved pharmacokinetics, enhanced therapeutic efficacy, and reduced off-target toxicity. Owing to their unique physicochemical properties, nanoparticles can be engineered to overcome biological barriers associated with tumor progression and facilitate precise delivery of therapeutic and diagnostic agents. This review comprehensively discusses the fundamental principles of cancer nanomedicine, including tumor biology, barriers to drug delivery, and critical design considerations for nanocarrier development. Various classes of nanomaterials, including polymeric nanoparticles, lipid-based systems, inorganic nanomaterials, and emerging biomimetic platforms, are examined with respect to their structural characteristics, therapeutic applications, and translational potential. Particular emphasis is placed on tumor-targeting strategies, encompassing passive, active, and microenvironment-responsive approaches, as well as on the development of smart stimuli-responsive nanocarriers capable of controlled, site-specific drug release. Furthermore, recent advances in nanotechnology-enabled chemotherapy, combination therapy, gene and RNA delivery, immuno-nanomedicine, and theranostic platforms are highlighted. The integration of diagnostic imaging and therapeutic functions within multifunctional nanocarriers has enabled real-time monitoring of treatment response and personalized cancer management. In addition, challenges associated with safety, toxicity, large-scale manufacturing, regulatory approval, and clinical translation are critically evaluated. Emerging innovations, including artificial intelligence-driven nanocarrier design, biomimetic nanomedicines, and precision oncology approaches, are also explored as future directions for the field. Overall, cancer nanomedicine has evolved from a simple drug-delivery concept into a multifunctional therapeutic platform integrating targeted therapy, molecular imaging, immunomodulation, gene therapy, and personalized medicine. Continued interdisciplinary collaboration and technological innovation are expected to accelerate the clinical translation of next-generation nanomedicines, ultimately improving treatment outcomes and advancing precision cancer care.

Keywords:

Cancer Nanomedicine; Nanoparticles; Targeted Drug Delivery; Smart Nanocarriers; Theranostics; Immuno-Nanomedicine; Gene Delivery; RNA Therapeutics; Precision Oncology; Tumor Microenvironment; Drug Delivery Systems; Cancer Therapy.

DOI: <https://doi.org/10.64062/JPGMB.Vol2.Issue4.2><https://jpgmb.com/1/issue/archive>

This is an Open Access article distributed under the terms of the Creative Commons Attribution (CC BY NC), which permits unrestricted use, distribution, and reproduction in any medium, as long as the original authors and source are cited. No permission is required from the authors or the publishers. (<https://creativecommons.org/licenses/by-nc/4.0/>)

1. Introduction

1.1 Cancer Nanomedicine

Cancer remains one of the leading causes of morbidity and mortality worldwide, accounting for millions of new diagnoses and deaths annually despite significant advances in diagnostic and therapeutic technologies¹. Conventional treatment modalities, including surgery, chemotherapy, and radiotherapy, have substantially improved patient outcomes; however, several critical limitations persist. These include non-specific systemic toxicity, inadequate tumor selectivity, multidrug resistance, unfavorable pharmacokinetic profiles, and insufficient drug accumulation at tumor sites². Consequently, there is a growing need for innovative therapeutic strategies capable of selectively targeting malignant tissues while minimizing damage to healthy cells.

Nanomedicine has emerged as a promising interdisciplinary field that integrates nanotechnology with oncology to improve cancer diagnosis, treatment, and monitoring³. Nanomedicine utilizes nanoscale materials, typically ranging from 1 to 200 nm in size, engineered to interact with biological systems at the molecular and cellular levels⁴. Owing to their unique physicochemical properties, including high surface-area-to-volume ratios, tunable surface chemistry, and controlled drug-release capabilities, nanoparticles serve as efficient carriers for therapeutic and diagnostic agents. These nanocarriers can protect encapsulated drugs from premature degradation, enhance aqueous solubility, improve pharmacokinetic behavior, prolong systemic circulation, and facilitate targeted drug delivery⁵.

The application of nanotechnology in oncology is particularly advantageous due to the distinctive characteristics of the tumor microenvironment. Solid tumors often exhibit abnormal vasculature, impaired lymphatic drainage, hypoxia, acidic pH conditions, and heterogeneous cellular composition⁶. These pathological features create opportunities for selective nanoparticle accumulation through passive targeting mechanisms, such as the enhanced permeability and retention (EPR) effect, while also enabling the development of active targeting strategies using tumor-specific ligands⁷. Consequently, nanomedicine can significantly improve the therapeutic index of anticancer agents by increasing drug concentration within tumors and reducing systemic exposure.

Over the past two decades, numerous nanomedicine-based formulations have progressed from preclinical development to clinical evaluation, demonstrating improved efficacy, enhanced bioavailability, and reduced toxicity compared with conventional therapies⁸. Beyond drug delivery, the scope of cancer nanomedicine has expanded to encompass diagnostic and theranostic applications. Theranostic nanoparticles integrate imaging and therapeutic

functionalities within a single platform, enabling real-time visualization of drug biodistribution, monitoring of therapeutic responses, and assessment of treatment resistance⁹. Such multifunctional systems support personalized treatment strategies by allowing clinicians to tailor therapeutic interventions according to individual patient characteristics and disease progression.

Recent advancements in nanotechnology have further facilitated the clinical translation of nucleic acid-based therapeutics, including small interfering RNA (siRNA), messenger RNA (mRNA), and gene-editing platforms, which would otherwise suffer from poor stability and rapid degradation under physiological conditions¹⁰. In addition, nanotechnology has become increasingly integrated with cancer immunotherapy, enhancing antigen delivery, improving immune-cell activation, and augmenting the therapeutic efficacy of immune checkpoint inhibitors and cancer vaccines¹¹. These developments have established nanomedicine as a critical component of next-generation precision oncology.

Despite remarkable progress, several challenges continue to hinder the widespread clinical implementation of cancer nanomedicines. Large-scale manufacturing remains a major obstacle due to difficulties in maintaining batch-to-batch reproducibility, physicochemical stability, and quality control during production¹². Furthermore, concerns regarding long-term safety, regulatory approval pathways, pharmacokinetic variability among patients, and the predictive value of preclinical models continue to complicate clinical translation¹³. Addressing these challenges requires a multidisciplinary approach involving tumor biology, materials science, pharmaceutical engineering, clinical oncology, and regulatory sciences.

This review explores the fundamental principles of cancer nanomedicine, including nanoparticle design strategies, mechanisms of tumor targeting, therapeutic and diagnostic applications, and emerging theranostic platforms. Additionally, it discusses current translational challenges and future opportunities for developing safer, more effective, and clinically relevant nanomedicine-based interventions for cancer management.

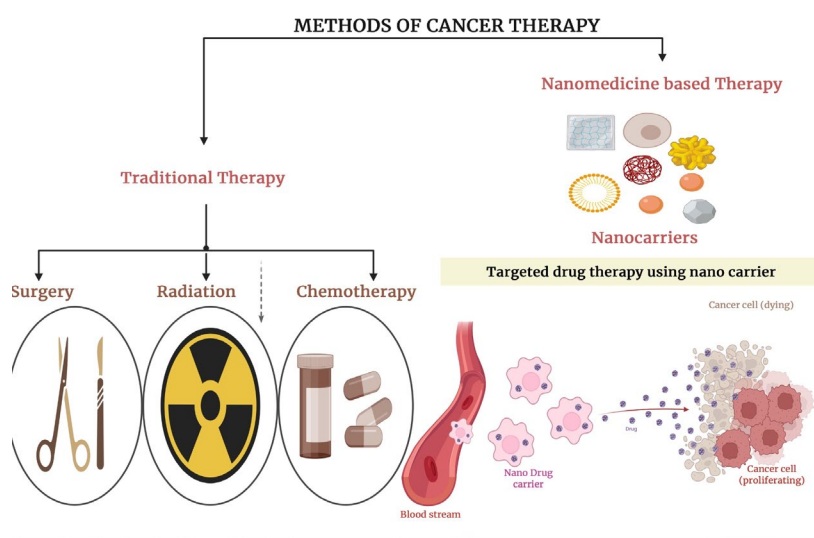


Figure 1: Conceptual comparison of traditional cancer therapies and nanomedicine-based targeted drug delivery.

1.2 Tumor Biology and Barriers to Drug Delivery

Cancer treatment remains challenging because of the highly heterogeneous nature of tumors and the complexity of the tumor microenvironment (TME). Unlike normal tissues, solid tumors develop within a pathological microenvironment characterized by abnormal angiogenesis, elevated interstitial fluid pressure (IFP), hypoxia, acidosis, dense extracellular matrix (ECM) deposition, and extensive cellular heterogeneity. These features collectively create substantial biological and physiological barriers that limit the effective delivery, penetration, and retention of therapeutic agents, ultimately contributing to treatment failure and drug resistance¹³.

One of the most prominent hallmarks of tumor biology is the presence of abnormal vasculature resulting from uncontrolled angiogenesis. Tumor blood vessels are structurally irregular, exhibiting heterogeneous diameters, discontinuous endothelial linings, poor pericyte coverage, and increased permeability. Although these abnormalities facilitate the extravasation of macromolecules and nanoparticles through the enhanced permeability and retention (EPR) effect, they also generate heterogeneous blood flow and poor tissue perfusion, leading to uneven drug distribution throughout the tumor mass^{14,15}. Consequently, certain tumor regions receive insufficient therapeutic exposure, thereby promoting the survival of resistant cancer cell populations.

Elevated interstitial fluid pressure represents another major obstacle to efficient drug delivery. Owing to dysfunctional lymphatic drainage and vascular leakage, fluid accumulates within tumor tissues, creating pressure gradients that oppose the convective transport of therapeutic agents from blood vessels into the tumor interstitium¹⁶. As a result, both conventional chemotherapeutics and nanoparticle-based formulations often remain concentrated in perivascular regions, while poorly perfused tumor cores receive inadequate treatment.

The extracellular matrix further restricts drug penetration within tumors. Compared with normal tissues, tumor-associated ECM is frequently enriched with collagen, fibronectin, hyaluronic acid, and other stromal components that increase tissue density and mechanical stiffness. This dense matrix acts as a physical barrier that impedes the diffusion of therapeutic molecules and nanocarriers while simultaneously influencing cellular signaling pathways associated with tumor progression and therapeutic resistance¹⁷. Cancer-associated fibroblasts actively remodel the ECM, further reinforcing transport barriers and reducing treatment efficacy.

Intratumoral heterogeneity presents an additional challenge for successful cancer therapy. Tumors consist of genetically, epigenetically, and phenotypically diverse cancer cell populations coexisting with stromal cells, immune cells, endothelial cells, and extracellular components. These distinct cellular populations often exhibit variable responses to therapeutic interventions, facilitating the emergence of drug-resistant subclones and limiting the effectiveness of single-target treatment strategies¹⁸. Consequently, achieving uniform drug distribution and therapeutic activity across all tumor compartments remains exceedingly difficult.

The tumor microenvironment also exhibits profound metabolic abnormalities. Poor vascular perfusion combined with high metabolic demand generates regions of hypoxia and acidosis, which significantly influence drug transport, cellular uptake, and nanoparticle stability¹⁹. Hypoxic conditions activate adaptive survival pathways that promote tumor aggressiveness and resistance to chemotherapy, radiotherapy, and immunotherapy. Similarly, acidic extracellular pH can alter drug ionization and compromise the stability and performance of nanocarrier systems.

At the cellular level, multiple biological barriers further hinder therapeutic efficacy. The plasma membrane serves as a selective barrier that restricts intracellular drug entry, while endosomal sequestration and efflux transporters such as P-glycoprotein (P-gp) actively reduce intracellular drug accumulation²⁰. These mechanisms contribute substantially to multidrug resistance and limit the therapeutic effectiveness of many anticancer agents. For nucleic acid-based therapeutics and biological drugs, successful treatment requires not only efficient cellular uptake but also effective escape from endosomal compartments before degradation occurs²¹.

1.3 Design Principles of Nanocarriers

The successful application of nanomedicine in oncology depends on the rational design of nanocarriers capable of overcoming systemic, tissue-level, and cellular barriers. Nanocarrier design requires careful optimization of physicochemical characteristics, including particle size, shape, surface properties, material composition, and drug-release behavior, to achieve favorable pharmacokinetics, enhanced tumor accumulation, efficient tissue penetration, and minimal off-target toxicity²².

Particle size is among the most critical determinants of nanocarrier biodistribution and therapeutic performance. Nanoparticles smaller than approximately 5–8 nm are rapidly eliminated through renal clearance, whereas particles larger than 200 nm are readily recognized and removed by the mononuclear phagocyte system (MPS), particularly in the liver and spleen²³. Nanocarriers within the optimal size range of approximately 20–150 nm generally exhibit prolonged circulation times and improved tumor accumulation via the EPR effect. However, the ideal particle size varies depending on tumor vascular permeability, stromal composition, and interstitial pressure, highlighting the need for tumor-specific nanocarrier engineering²⁴.

Nanoparticle shape and mechanical properties also significantly influence biological behavior. While most clinically approved nanomedicines utilize spherical particles, alternative geometries such as rod-shaped nanoparticles, filamentous micelles, and disc-shaped structures have demonstrated prolonged circulation and reduced macrophage uptake in preclinical studies^{25,26}. Similarly, particle rigidity affects biodistribution and tumor penetration. Soft and deformable nanocarriers often display superior extravasation and deeper penetration into tumor tissues compared with rigid nanoparticles²⁷.

Surface characteristics play a crucial role in determining interactions between nanocarriers and biological systems. Positively charged nanoparticles typically exhibit enhanced cellular uptake but may also promote protein adsorption, complement activation, and systemic toxicity²⁸. In

contrast, neutral or slightly negatively charged particles generally demonstrate improved circulation stability and reduced immune recognition. Surface modification with hydrophilic polymers such as polyethylene glycol (PEG) has been widely employed to extend circulation time and reduce opsonization. However, repeated administration of PEGylated nanoparticles may induce anti-PEG antibodies, resulting in accelerated blood clearance and reduced therapeutic efficacy²⁹. Consequently, alternative stealth coatings and biomimetic surface engineering strategies are being actively investigated.

Material composition further influences drug-loading capacity, degradation kinetics, safety, and clinical translatability. Lipid-based and polymeric nanocarriers offer excellent biocompatibility and tunable release profiles, whereas inorganic nanoparticles provide unique imaging capabilities and responsiveness to external stimuli³⁰. Importantly, biodegradability and metabolite safety remain essential considerations, as non-degradable materials may accumulate in healthy tissues and raise long-term toxicity concerns.

Drug-loading strategies and release mechanisms constitute additional critical design parameters. Effective nanocarriers must achieve high drug encapsulation efficiency while minimizing premature drug leakage during circulation. Controlled drug release can be achieved through diffusion, matrix degradation, or stimuli-responsive mechanisms triggered by pH, enzymes, redox conditions, temperature, or external energy sources³¹. Furthermore, for nucleic acid and protein therapeutics, successful endosomal escape remains a major challenge. To address this limitation, advanced nanocarrier systems increasingly incorporate pH-responsive, membrane-disruptive, or redox-sensitive components that facilitate cytosolic delivery and enhance therapeutic efficacy³².

Table :1: Design Parameters of Nanocarriers and Their Biological Implications

Parameter	Characteristics	Biological Impact	Reference
Nanoparticle size	20-150nm	increased tumor accumulation, short half-life, avoid renal clearance	23
NanoParticle shape	rod-shaped, filamentous micelles, disk shape	Alteration in margination, increase in macrophage uptake, increase internalization of cell	26
Surface charge	Neutral to slight negative	Low protein absorption to increase circulation stability	29
Material composition	Polymeric, lipid-based	Low protein opsonization to increase circulation time	30

Surface chemistry	polyethylene glycosylation	Drug loading determination, stimulus responsiveness	29
Drug loading	Encapsulation, adsorption, covalent conjugation	Drug cargo protection, enhance solubility and stability	31
Drug release	Diffusion-controlled, degradation-mediated, stimuli-responsive	Temporal control of drug action	30
Intracellular trafficking	pH-responsive or membrane-disruptive elements	Enhance cytosolic delivery	32

1.4 Polymeric and Lipid-Based Nanoparticles

Polymeric and lipid-based nanoparticles represent the most extensively investigated nanocarrier platforms in cancer nanomedicine owing to their excellent biocompatibility, structural versatility, and tunable drug delivery characteristics³³. These systems have demonstrated significant potential for overcoming the pharmacokinetic and pharmacodynamic limitations associated with conventional anticancer therapies by enabling controlled drug release, enhancing tumor accumulation, and minimizing systemic toxicity. Their adaptable physicochemical properties allow for the optimization of therapeutic efficacy while improving the safety profile of anticancer agents.

Among polymeric nanocarriers, biodegradable polymers such as poly(lactic-co-glycolic acid) (PLGA), poly(lactic acid) (PLA), polycaprolactone (PCL), chitosan, and alginate have received considerable attention for cancer drug delivery applications³⁴. The chemical flexibility of these materials enables precise control over particle size, surface characteristics, drug-loading capacity, and degradation kinetics. Therapeutic agents may be physically encapsulated within the polymeric matrix or chemically conjugated to polymer backbones, allowing sustained and predictable drug release through mechanisms involving polymer degradation and diffusion. Polymeric nanoparticles have proven effective for the delivery of hydrophobic chemotherapeutic agents, proteins, peptides, and nucleic acid therapeutics. Furthermore, surface functionalization strategies can impart active targeting capabilities and stealth properties, thereby improving tumor selectivity and prolonging systemic circulation. Despite these advantages, challenges including burst drug release, formulation complexity, manufacturing scalability, and batch-to-batch reproducibility continue to limit broader clinical translation³⁵.

Polymeric micelles constitute a specialized class of self-assembled nanocarriers formed from amphiphilic block copolymers. These nanostructures possess a hydrophobic core surrounded by a hydrophilic corona, making them particularly suitable for the solubilization and delivery

of poorly water-soluble anticancer drugs³⁶. Encapsulation within the hydrophobic core enhances drug stability, improves aqueous solubility, and prolongs circulation time relative to free drug molecules. Several micellar formulations have demonstrated encouraging outcomes in preclinical and clinical studies. Nevertheless, micellar systems remain susceptible to dilution-induced dissociation following intravenous administration, potentially leading to premature drug leakage and reduced therapeutic efficiency³⁷.

Lipid-based nanoparticles have emerged as one of the most clinically successful nanomedicine platforms. This category includes liposomes, solid lipid nanoparticles (SLNs), nanostructured lipid carriers (NLCs), and lipid nanoparticles (LNPs), all of which exhibit favorable biocompatibility and versatile drug-loading capabilities³⁸. Liposomes are composed of phospholipid bilayers enclosing aqueous compartments capable of accommodating both hydrophilic and hydrophobic therapeutic agents. Due to their biomimetic structure and excellent safety profile, liposomal formulations have been widely adopted for cancer therapy. Surface modification through polyethylene glycol (PEG) conjugation further enhances circulation half-life and reduces immune system recognition, thereby increasing tumor accumulation via passive targeting mechanisms³⁹. Several liposomal anticancer formulations have received regulatory approval, highlighting the successful clinical translation of lipid-based nanomedicines.

More recently, lipid nanoparticles have gained considerable attention as efficient carriers for nucleic acid therapeutics, including small interfering RNA (siRNA), messenger RNA (mRNA), and gene-editing systems⁴⁰. A critical component of these systems is the incorporation of ionizable lipids, which facilitate endosomal escape and promote efficient intracellular delivery of genetic cargo. Consequently, lipid nanoparticles have become attractive platforms for gene-based cancer therapies and precision medicine applications. Despite their therapeutic promise, lipid-based systems are not without limitations. Potential challenges include infusion-related adverse reactions, hepatic accumulation, immunogenicity, and formulation-dependent toxicity. Therefore, careful selection of lipid components and optimization of manufacturing processes remain essential for maximizing therapeutic performance and clinical safety⁴¹.

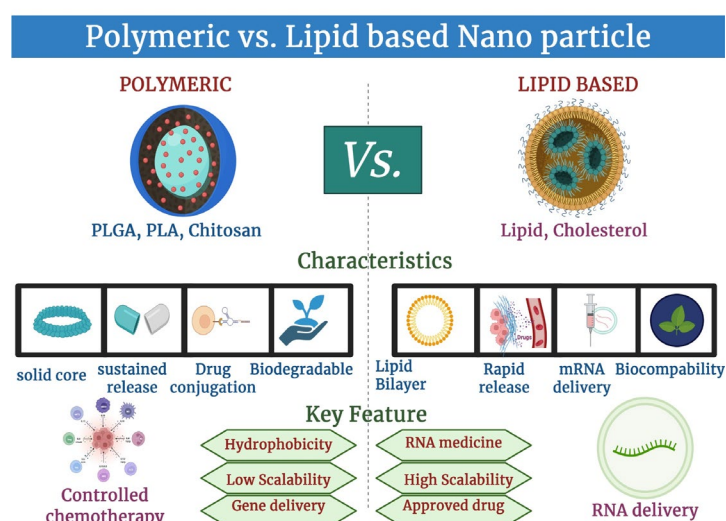


Figure 2: Comparison of polymeric and lipid-based nanoparticle platforms used in cancer nanomedicine. Created in <https://BioRender.com>

1.5 Inorganic and Emerging Nanomaterials

Inorganic nanomaterials have emerged as highly versatile platforms in cancer nanomedicine due to their unique physicochemical properties that extend beyond the capabilities of conventional polymeric and lipid-based systems. These nanomaterials possess tunable optical, magnetic, electrical, and catalytic characteristics, enabling their application in drug delivery, molecular imaging, biosensing, and externally triggered therapeutic interventions⁴².

Among inorganic nanomaterials, gold nanoparticles (AuNPs) have attracted considerable attention owing to their remarkable surface plasmon resonance properties. Their optical behavior can be precisely tuned by controlling particle size and morphology, making them highly suitable for photothermal therapy, image-guided treatment, radiosensitization, and multimodal cancer theranostics. Furthermore, gold nanoparticles can be readily functionalized with targeting ligands, therapeutic agents, and surface polymers, allowing simultaneous diagnostic and therapeutic applications. Despite these advantages, concerns regarding long-term tissue accumulation, limited biodegradability, and potential cytotoxicity necessitate careful optimization of particle size, surface chemistry, and dosing strategies⁴³.

Iron oxide nanoparticles represent another important class of inorganic nanocarriers with significant applications in magnetic resonance imaging (MRI), magnetic targeting, and hyperthermia-based cancer therapy. Their superparamagnetic properties enable tumor localization using external magnetic fields while facilitating image-guided therapeutic interventions. Exposure to alternating magnetic fields can generate localized hyperthermia, inducing selective tumor cell destruction. Although preclinical studies have demonstrated encouraging outcomes, challenges including nanoparticle aggregation, nonspecific organ accumulation, and dose-dependent toxicity continue to hinder widespread clinical translation⁴⁴.

Silica-based nanomaterials, particularly mesoporous silica nanoparticles (MSNs), have gained considerable interest because of their exceptionally high surface area, tunable pore architecture, and excellent physicochemical stability. These features enable high drug-loading efficiency and precise control over drug-release kinetics. Surface modification with targeting ligands and stimuli-responsive gatekeepers further enhances their therapeutic performance. Nevertheless, concerns regarding slow biodegradation, prolonged tissue retention, and long-term biocompatibility remain significant barriers to clinical application⁴⁴.

Carbon-based nanomaterials, including carbon nanotubes, graphene derivatives, graphene oxide, and carbon quantum dots, have also been extensively investigated for cancer therapy. These materials exhibit unique mechanical strength, electrical conductivity, photothermal conversion efficiency, and large surface areas suitable for drug loading. In addition to serving as drug and nucleic acid carriers, carbon nanomaterials have shown promise in cancer imaging, photothermal ablation, and biosensing applications. However, their intrinsic hydrophobicity, potential immunogenicity, and long-term persistence within biological tissues necessitate

extensive surface engineering and comprehensive toxicity evaluation before clinical implementation⁴⁵.

Recent advances have further expanded the field toward emerging nanomaterials, including biomimetic nanocarriers, hybrid organic-inorganic nanocomposites, and multifunctional stimuli-responsive systems. Biomimetic nanoparticles utilize natural biological membranes derived from erythrocytes, platelets, leukocytes, or cancer cells to evade immune surveillance and enhance tumor targeting. Hybrid nanomaterials combine the advantageous properties of multiple material classes, enabling simultaneous imaging, targeted drug delivery, and therapeutic intervention within a single platform. Collectively, these emerging technologies significantly broaden the functional capabilities of cancer nanomedicine while offering opportunities for precision oncology and personalized treatment strategies⁴⁶. Nevertheless, challenges associated with biodegradability, safety, manufacturing scalability, and regulatory approval must be addressed to facilitate successful clinical translation⁴⁷.

1.6 Targeting Strategies in Nanodrug Delivery

Targeting strategies constitute a fundamental component of cancer nanomedicine, aiming to maximize therapeutic efficacy while minimizing systemic toxicity. Unlike conventional chemotherapeutic agents, which distribute non-selectively throughout the body, nanocarrier-based systems can be rationally engineered to preferentially accumulate within tumors and selectively interact with malignant cells⁴⁸. Current nanomedicine approaches primarily employ passive targeting, active targeting, and biological or microenvironment-responsive targeting mechanisms⁴⁹.

Passive targeting exploits the unique pathophysiological characteristics of solid tumors, particularly the enhanced permeability and retention (EPR) effect. Tumor vasculature is typically characterized by abnormal endothelial architecture, large fenestrations, and impaired lymphatic drainage, allowing nanoparticles of appropriate size to extravasate and accumulate within tumor tissues more efficiently than in healthy organs⁵⁰. However, reliance solely on passive targeting often results in variable therapeutic outcomes because tumor vascular permeability, stromal density, and interstitial fluid pressure differ substantially among patients and tumor types. Consequently, nanoparticle distribution within tumors remains highly heterogeneous, limiting treatment effectiveness^{49, 50}.

Active targeting seeks to improve cellular specificity through surface functionalization of nanocarriers with ligands capable of recognizing overexpressed receptors on cancer cells or tumor-associated tissues. Common targeting ligands include monoclonal antibodies, peptides, aptamers, carbohydrates, and small molecules such as folic acid. Upon ligand-receptor interaction, receptor-mediated endocytosis facilitates enhanced intracellular uptake and retention of therapeutic payloads within target cells⁵¹. Although active targeting does not necessarily increase total tumor accumulation, it significantly improves cellular internalization and therapeutic selectivity. Nevertheless, challenges such as receptor heterogeneity, variable receptor expression, ligand density optimization, and potential immunogenic responses continue to affect clinical performance.

Biological and microenvironment-responsive targeting strategies represent a more advanced approach that exploits unique biological features of tumors, including altered metabolism, immune interactions, hypoxia, acidity, and enzymatic activity. Biomimetic nanocarriers coated with natural cell membranes can evade immune clearance and leverage endogenous biological recognition pathways to achieve prolonged circulation and enhanced tumor homing⁵². By mimicking native cellular functions, these systems often exhibit improved biocompatibility and targeting efficiency without requiring synthetic targeting ligands⁵³. Such approaches are increasingly being integrated with conventional targeting strategies to improve therapeutic precision and overcome biological barriers associated with cancer drug delivery.

1.7 Smart and Functional Nanomedicine Platforms for Cancer Therapy

The development of smart and multifunctional nanomedicine platforms has transformed the landscape of cancer therapy by enabling dynamic interactions between nanocarriers and the tumor microenvironment. Unlike conventional delivery systems that rely primarily on passive accumulation or receptor-mediated targeting, smart nanomedicines possess responsive functionalities that facilitate controlled drug release, enhanced intracellular delivery, and synergistic therapeutic effects⁵⁴. These advanced systems are specifically designed to improve treatment precision, overcome therapeutic resistance, and minimize off-target toxicity.

1.7.1 Stimuli-Responsive and Smart Nanocarriers

Stimuli-responsive nanocarriers are engineered to release therapeutic payloads selectively in response to specific endogenous or exogenous triggers. These systems exploit physiological differences between tumor tissues and normal tissues to achieve site-specific drug delivery and controlled therapeutic activation.

Among endogenous stimuli, acidic pH, elevated intracellular glutathione concentrations, reactive oxygen species, and tumor-associated enzymes are the most extensively investigated triggers. The extracellular environment of many tumors exhibits mild acidity, while intracellular endosomal and lysosomal compartments possess even lower pH values. pH-responsive nanocarriers utilize protonation-induced structural destabilization to trigger selective drug release within tumor tissues or intracellular compartments⁵⁵. Similarly, redox-responsive systems exploit the elevated glutathione concentrations found in cancer cells, enabling intracellular cleavage of disulfide bonds and subsequent release of therapeutic agents. Enzyme-responsive nanocarriers further enhance selectivity by utilizing tumor-associated proteases, such as matrix metalloproteinases and cathepsins, to activate drug release specifically within the tumor microenvironment³².

Exogenous stimuli, including light, ultrasound, heat, and magnetic fields, provide an additional level of control over therapeutic activation. These externally triggered systems allow clinicians to precisely regulate the timing and location of drug release, often in conjunction with imaging-guided treatment strategies. Photothermal and photodynamic nanomedicines, for example, utilize near-infrared light to activate therapeutic responses selectively within tumor tissues.

Similarly, magnetic and ultrasound-responsive platforms enable non-invasive modulation of drug release and therapeutic efficacy⁵⁶. Despite their considerable promise, the clinical implementation of these technologies remains constrained by factors such as limited tissue penetration depth, equipment requirements, and challenges associated with large-scale manufacturing and standardization.

Collectively, stimuli-responsive nanocarriers represent a major advancement toward precision oncology by providing spatiotemporal control over therapeutic delivery, reducing systemic exposure, and enhancing treatment outcomes in diverse cancer types.

1.8 Conclusion

Cancer nanomedicine has advanced from a conceptual strategy for improving drug delivery into a broad and multifunctional discipline that integrates nanocarrier engineering, tumor biology, targeting science, and theranostic innovation. The rational design of nanocarriers, guided by careful control of particle size, shape, surface chemistry, and material composition, has enabled substantial improvements in circulation time, tumor accumulation, and controlled drug release. Polymeric, lipid-based, and inorganic nanomaterials each offer distinct advantages, and their continued refinement, together with the emergence of biomimetic and hybrid platforms, has expanded the functional scope of nanomedicine well beyond simple drug encapsulation.

Targeting strategies, spanning passive accumulation, active receptor-mediated recognition, and biological or microenvironment-responsive approaches, have collectively improved the precision with which therapeutic agents reach malignant tissue while sparing healthy organs. The development of stimuli-responsive and smart nanocarrier systems further reinforces this progress by allowing spatiotemporal control over drug release, thereby enhancing therapeutic index and reducing systemic toxicity. Nonetheless, translating these advances into routine clinical practice remains constrained by challenges in large-scale manufacturing, batch-to-batch reproducibility, long-term biocompatibility, and the complexity of regulatory approval pathways.

Looking forward, continued interdisciplinary collaboration among materials scientists, tumor biologists, clinicians, and regulatory bodies will be essential to bridge the gap between preclinical promise and clinical translation. Emerging directions, including artificial intelligence-guided nanocarrier design, biomimetic engineering, and integrated theranostic platforms, hold considerable potential to further personalize cancer treatment. With sustained innovation and rigorous evaluation of safety and efficacy, cancer nanomedicine is well positioned to become an increasingly central pillar of precision oncology, ultimately improving therapeutic outcomes and quality of life for cancer patients.

References:

1. Farhood, B., Najafi, M., & Mortezaee, K. (2019). CD8+ cytotoxic T lymphocytes in cancer immunotherapy: A review. *Journal of Cellular Physiology*, 234(6), 8509–8521. <https://doi.org/10.1002/jcp.27782>
2. Ferris, R. L., Blumenschein, G., Fayette, J., Guigay, J., Colevas, A. D., Licitra, L., Harrington, K., Kasper, S., Vokes, E. E., Even, C., Worden, F., Saba, N. F., Iglesias Docampo, L. C., Haddad, R., Rordorf, T., Kiyota, N., Tahara, M., Monga, M., Lynch, M., Geese, W. J., Kopit, J., Shaw, J. W., & Gillison, M. L. (2016). Nivolumab for recurrent squamous-cell carcinoma of the head and neck. *New England Journal of Medicine*, 375(19), 1856–1867. <https://doi.org/10.1056/NEJMoa1602252>
3. Bellmunt, J., de Wit, R., Vaughn, D. J., Fradet, Y., Lee, J. L., Fong, L., Vogelzang, N. J., Climent, M. A., Petrylak, D. P., Choueiri, T. K., Necchi, A., Gerritsen, W., Gurney, H., Quinn, D. I., Culine, S., Sternberg, C. N., Mai, Y., Poehlein, C. H., Perini, R. F., & Bajorin, D. F. (2017). Pembrolizumab as second-line therapy for advanced urothelial carcinoma. *New England Journal of Medicine*, 376(11), 1015–1026. <https://doi.org/10.1056/NEJMoa1613683>
4. Chung, C. (2018). Restoring the switch for cancer cell death: Targeting the apoptosis signaling pathway. *American Journal of Health-System Pharmacy*, 75(13), 945–952. <https://doi.org/10.2146/ajhp170607>
5. Saito, H., Takaya, S., Osaki, T., & Ikeguchi, M. (2013). Increased apoptosis and elevated Fas expression in circulating natural killer cells in gastric cancer patients. *Gastric Cancer*, 16(4), 473–479. <https://doi.org/10.1007/s10120-012-0210-1>
6. Mohme, M., Riethdorf, S., & Pantel, K. (2017). Circulating and disseminated tumour cells: Mechanisms of immune surveillance and escape. *Nature Reviews Clinical Oncology*, 14(3), 155–167. <https://doi.org/10.1038/nrclinonc.2016.144>
7. Motz, G. T., Santoro, S. P., Wang, L. P., Garrabrant, T., Lastra, R. R., Hagemann, I. S., Lal, P., Feldman, M. D., Benencia, F., & Coukos, G. (2014). Tumor endothelium FasL establishes a selective immune barrier promoting tolerance in tumors. *Nature Medicine*, 20(6), 607–615. <https://doi.org/10.1038/nm.3541>
8. Wang, Y. H., & Scadden, D. T. (2015). Harnessing the apoptotic programs in cancer stem-like cells. *EMBO Reports*, 16(9), 1084–1098. <https://doi.org/10.15252/embr.201439675>
9. Steinbichler, T. B., Dudás, J., Skvortsov, S., Ganswindt, U., Riechelmann, H., & Skvortsova, I. I. (2018). Therapy resistance mediated by cancer stem cells. *Seminars in Cancer Biology*, 53, 156–167. <https://doi.org/10.1016/j.semcancer.2018.11.006>
10. Phi, L. T. H., Sari, I. N., Yang, Y. G., Lee, S. H., Jun, N., Kim, K. S., Lee, Y. K., & Kwon, H. Y. (2018). Cancer stem cells (CSCs) in drug resistance and their therapeutic implications in cancer treatment. *Stem Cells International*, 2018, 5416923. <https://doi.org/10.1155/2018/5416923>
11. Li, Y., Wang, Z., Ajani, J. A., & Song, S. (2021). Drug resistance and cancer stem cells. *Cell Communication and Signaling*, 19(1), 19. <https://doi.org/10.1186/s12964-020-00627-5>
12. Costello, R. T., Mallet, F., Gaugler, B., Sainty, D., Arnoulet, C., Gastaut, J. A., & Olive, D. (2000). Human acute myeloid leukemia CD34+/CD38– progenitor cells have

- decreased sensitivity to chemotherapy and Fas-induced apoptosis, reduced immunogenicity, and impaired dendritic cell transformation capacities. *Cancer Research*, 60(16), 4403–4411.
13. Peitzsch, C., Kurth, I., Kunz-Schughart, L., Baumann, M., & Dubrovskaya, A. (2013). Discovery of the cancer stem cell-related determinants of radioresistance. *Radiotherapy and Oncology*, 108(3), 378–387. <https://doi.org/10.1016/j.radonc.2013.06.003>
 14. Haga, R. B., & Ridley, A. J. (2016). Rho GTPases: Regulation and roles in cancer cell biology. *Small GTPases*, 7(4), 207–221. <https://doi.org/10.1080/21541248.2016.1232583>
 15. Green, D. R., & Fitzgerald, P. (2016). Just so stories about the evolution of apoptosis. *Current Biology*, 26(13), R620–R627. <https://doi.org/10.1016/j.cub.2016.05.023>
 16. Yang, J., Chai, X. Q., Zhao, X. X., & Li, X. (2017). Comparative genomics revealed the origin and evolution of autophagy pathway. *Journal of Systematics and Evolution*, 55(1), 71–82. <https://doi.org/10.1111/jse.12212>
 17. Mariño, G., Niso-Santano, M., Baehrecke, E. H., & Kroemer, G. (2014). Self-consumption: The interplay of autophagy and apoptosis. *Nature Reviews Molecular Cell Biology*, 15(2), 81–94. <https://doi.org/10.1038/nrm3735>
 18. Germain, M., Mathai, J. P., & Shore, G. C. (2002). BH3-only BIK functions at the endoplasmic reticulum to stimulate cytochrome c release from mitochondria. *Journal of Biological Chemistry*, 277(20), 18053–18060. <https://doi.org/10.1074/jbc.M201235200>
 19. Amr, S., Heisey, C., Zhang, M., Xia, X. J., Shows, K. H., Ajlouni, K., Pandya, A., Satin, L. S., El-Shanti, H., & Shiang, R. (2007). A homozygous mutation in a novel zinc-finger protein, ERIS, is responsible for Wolfram syndrome 2. *American Journal of Human Genetics*, 81(4), 673–683. <https://doi.org/10.1086/520961>
 20. Zhu, W., Cowie, A., Wasfy, G. W., Penn, L. Z., Leber, B., & Andrews, D. W. (1996). Bcl-2 mutants with restricted subcellular location reveal spatially distinct pathways for apoptosis in different cell types. *The EMBO Journal*, 15(16), 4130–4141.
 21. Chang, N. C., Nguyen, M., Germain, M., & Shore, G. C. (2010). Antagonism of Beclin 1-dependent autophagy by BCL-2 at the endoplasmic reticulum requires NAF-1. *The EMBO Journal*, 29(3), 606–618. <https://doi.org/10.1038/emboj.2009.369>
 22. Baehrecke, E. H. (2005). Autophagy: Dual roles in life and death? *Nature Reviews Molecular Cell Biology*, 6(6), 505–510. <https://doi.org/10.1038/nrm1666>
 23. Pickrell, A. M., & Youle, R. J. (2015). The roles of PINK1, Parkin, and mitochondrial fidelity in Parkinson's disease. *Neuron*, 85(2), 257–273. (Note: The DOI provided in the source appears incorrect and should be verified.)
 24. Maiuri, M. C., Zalckvar, E., Kimchi, A., & Kroemer, G. (2007). Self-eating and self-killing: Crosstalk between autophagy and apoptosis. *Nature Reviews Molecular Cell Biology*, 8(9), 741–752. <https://doi.org/10.1038/nrm2239>
 25. Sipos, F., Firneisz, G., & Múzes, G. (2016). Therapeutic aspects of c-MYC signaling in inflammatory and cancerous colonic diseases. *World Journal of Gastroenterology*, 22(35), 7938–7950. <https://doi.org/10.3748/wjg.v22.i35.7938>
 26. Manié, S. N., Lebeau, J., & Chevet, E. (2014). Cellular mechanisms of endoplasmic reticulum stress signaling in health and disease. *American Journal of Physiology-Cell Physiology*, 307(10), C901–C907. <https://doi.org/10.1152/ajpcell.00292.2014>

27. Butler, D. E., Marlein, C., Walker, H. F., Frame, F. M., Mann, V. M., Simms, M. S., Davies, B. R., Collins, A. T., & Maitland, N. J. (2017). Inhibition of the PI3K/AKT/mTOR pathway activates autophagy and compensatory Ras/Raf/MEK/ERK signalling in prostate cancer. *Oncotarget*, 8(34), 56698–56713. <https://doi.org/10.18632/oncotarget.18082>
28. González-Polo, R. A., Boya, P., Pauleau, A. L., Jalil, A., Larochette, N., Souquère, S., Eskelinen, E. L., Pierron, G., Saftig, P., & Kroemer, G. (2005). The apoptosis/autophagy paradox: Autophagic vacuolization before apoptotic death. *Journal of Cell Science*, 118(14), 3091–3102. <https://doi.org/10.1242/jcs.02447>
29. Zhao, Z., Xia, G., Li, N., Su, R., Chen, X., & Zhong, L. (2018). Autophagy inhibition promotes bevacizumab-induced apoptosis and proliferation inhibition in colorectal cancer cells. *Journal of Cancer*, 9(18), 3407–3416. <https://doi.org/10.7150/jca.24201>
30. Das, D. N., Naik, P. P., Mukhopadhyay, S., Panda, P. K., Sinha, N., Meher, B. R., & Bhutia, S. K. (2017). Elimination of dysfunctional mitochondria through mitophagy suppresses benzo[a]pyrene-induced apoptosis. *Free Radical Biology and Medicine*, 112, 452–463. <https://doi.org/10.1016/j.freeradbiomed.2017.08.020>
31. Grasso, D., Ropolo, A., Lo Re, A., Ré, V., Boggio, M. I., Molejón, J. L., Iovanna, J. L., Gonzalez, C. D., Urrutia, R., & Vaccaro, M. I. (2011). Zymophagy, a novel selective autophagy pathway mediated by VMP1-USP9x-p62, prevents pancreatic cell death. *Journal of Biological Chemistry*, 286(10), 8308–8324. <https://doi.org/10.1074/jbc.M110.197301>
32. Hou, W., Han, J., Lu, C., Goldstein, L. A., & Rabinowich, H. (2010). Autophagic degradation of active caspase-8: A crosstalk mechanism between autophagy and apoptosis. *Autophagy*, 6(7), 891–900. <https://doi.org/10.4161/auto.6.7.13038>
33. Pagliarini, V., Wirawan, E., Romagnoli, A., Ciccocanti, F., Lisi, G., Lippens, S., Cecconi, F., Fimia, G. M., Vandenabeele, P., Corazzari, M., & Pientini, M. (2012). Proteolysis of Ambra1 during apoptosis has a role in the inhibition of the autophagic pro-survival response. *Cell Death and Differentiation*, 19(9), 1495–1504. <https://doi.org/10.1038/cdd.2012.27>
34. Lamy, L., Ngo, V. N., Emre, N. C. T., Shaffer, A. L., Yang, Y., Tian, E., Nair, V., Kruhlak, M. J., Zingone, A., Landgren, O., & Staudt, L. M. (2013). Control of autophagic cell death by caspase-10 in multiple myeloma. *Cancer Cell*, 23(4), 435–449. <https://doi.org/10.1016/j.ccr.2013.02.017>
35. Pyo, J. O., Jang, M. H., Kwon, Y. K., Lee, H. J., Jun, J. I., Woo, H. N., Cho, D. H., Choi, B. Y., Lee, H., Kim, J. H., Mizushima, N., Ohsumi, Y., & Jung, Y. K. (2005). Essential roles of Atg5 and FADD in autophagic cell death: Dissection of autophagic cell death into vacuole formation and cell death. *Journal of Biological Chemistry*, 280(21), 20722–20729. <https://doi.org/10.1074/jbc.M413934200>
36. Ye, X., Zhou, X. J., & Zhang, H. (2018). Exploring the role of autophagy-related gene 5 (ATG5) yields important insights into autophagy in autoimmune/autoinflammatory diseases. *Frontiers in Immunology*, 9, 2334. <https://doi.org/10.3389/fimmu.2018.02334>
37. Nian, H., & Ma, B. (2021). Calpain-calpastatin system and cancer progression. *Biological Reviews*, 96(3), 961–975. <https://doi.org/10.1111/brv.12686>
38. Zhang, J., Zhang, S., Shi, Q., Allen, T. D., You, F., & Yang, D. (2020). The anti-apoptotic proteins Bcl-2 and Bcl-xL suppress Beclin 1/Atg6-mediated lethal autophagy

- in polyploid cells. *Experimental Cell Research*, 394(2), 112112. <https://doi.org/10.1016/j.yexcr.2020.112112>
39. Gordy, C., & He, Y. W. (2012). The crosstalk between autophagy and apoptosis: Where does this lead? *Protein & Cell*, 3(1), 17–27. <https://doi.org/10.1007/s13238-011-1127-x>
 40. Heras-Sandoval, D., Pérez-Rojas, J. M., Hernández-Damián, J., & Pedraza-Chaverri, J. (2014). The role of PI3K/AKT/mTOR pathway in the modulation of autophagy and the clearance of protein aggregates in neurodegeneration. *Cellular Signalling*, 26(12), 2694–2701. <https://doi.org/10.1016/j.cellsig.2014.08.019>
 41. Xie, W., Wulin, H., Shao, G., Wei, L., Qi, R., Ma, B., Chen, N., & Shi, R. (2020). Polygalasaponin F inhibits neuronal apoptosis induced by oxygen-glucose deprivation and reoxygenation through the PI3K/Akt pathway. *Basic & Clinical Pharmacology & Toxicology*, 127(3), 196–204. <https://doi.org/10.1111/bcpt.13408>
 42. Safa, A. R. (2013). Roles of c-FLIP in apoptosis, necroptosis, and autophagy. *Journal of Carcinogenesis & Mutagenesis, Suppl 6*, 003. <https://doi.org/10.4172/2157-2518.S6-003>
 43. Singh, P., Ravanani, P., & Talwar, P. (2016). Death-associated protein kinase 1 (DAPK1): A regulator of apoptosis and autophagy. *Frontiers in Molecular Neuroscience*, 9, 46. <https://doi.org/10.3389/fnmol.2016.00046>
 44. Elbadawy, M., Usui, T., Yamawaki, H., & Sasaki, K. (2018). Novel functions of death-associated protein kinases through mitogen-activated protein kinase-related signals. *International Journal of Molecular Sciences*, 19(10), 3031. <https://doi.org/10.3390/ijms19103031>
 45. Yu, L., Wan, F., Dutta, S., Welsh, S., Liu, Z. H., Freundt, E., Baehrecke, E. H., & Lenardo, M. (2006). Autophagic programmed cell death by selective catalase degradation. *Proceedings of the National Academy of Sciences of the United States of America*, 103(13), 4952–4957. <https://doi.org/10.1073/pnas.0511288103>
 46. Bonapace, L., Bornhauser, B. C., Schmitz, M., Cario, G., Ziegler, U., Niggli, F. K., Schäfer, B. W., Schrappe, M., Stanulla, M., & Bourquin, J. P. (2010). Induction of autophagy-dependent necroptosis is required for childhood acute lymphoblastic leukemia cells to overcome glucocorticoid resistance. *Journal of Clinical Investigation*, 120(4), 1310–1323. <https://doi.org/10.1172/JCI39987>
 47. Yao, Z., Zhang, P., Guo, H., Shi, J., Liu, S., Liu, Y., & Zheng, D. (2015). RIP1 modulates death receptor-mediated apoptosis and autophagy in macrophages. *Molecular Oncology*, 9(4), 806–817. <https://doi.org/10.1016/j.molonc.2014.12.004>
 48. Verzella, D., Pescatore, A., Capece, D., Vecchiotti, D., Ursini, M. V., Franzoso, G., Alesse, E., & Zazzeroni, F. (2020). Life, death, and autophagy in cancer: NF-κB turns up everywhere. *Cell Death & Disease*, 11(3), 210. <https://doi.org/10.1038/s41419-020-2399-y>
 49. Blaser, H., Dostert, C., Mak, T. W., & Brenner, D. (2016). TNF and ROS crosstalk in inflammation. *Trends in Cell Biology*, 26(4), 249–261. <https://doi.org/10.1016/j.tcb.2015.12.002>
 50. Ryan, K. M., Ernst, M. K., Rice, N. R., & Vousden, K. H. (2000). Role of NF-κB in p53-mediated programmed cell death. *Nature*, 404(6780), 892–897. <https://doi.org/10.1038/35009130>

51. Khandelwal, N., Simpson, J., Taylor, G., Rafique, S., Whitehouse, A., Hiscox, J., & Stark, L. A. (2011). Nucleolar NF- κ B/RelA mediates apoptosis by causing cytoplasmic relocalization of nucleophosmin. *Cell Death & Differentiation*, 18(12), 1889–1903. <https://doi.org/10.1038/cdd.2011.79>
52. Kong, P., Zhu, X., Geng, Q., Xia, L., Sun, X., Chen, Y., Li, W., Zhou, Z., Zhan, Y., & Xu, D. (2017). The microRNA-423-3p-Bim axis promotes cancer progression and activates oncogenic autophagy in gastric cancer. *Molecular Therapy*, 25(4), 1027–1037. <https://doi.org/10.1016/j.ymthe.2017.01.013>
53. Su, B., Wang, X. T., Sun, Y. H., Long, M. Y., Zheng, J., Wu, W. H., & Li, L. (2020). Ischemia/hypoxia inhibits cardiomyocyte autophagy and promotes apoptosis via the Egr-1/Bim/Beclin-1 pathway. *Journal of Geriatric Cardiology*, 17(5), 284–293. <https://doi.org/10.11909/j.issn.1671-5411.2020.05.004>
54. Yang, D., Okamura, H., Teramachi, J., & Haneji, T. (2016). Histone demethylase Jmjd3 regulates osteoblast apoptosis through targeting anti-apoptotic protein Bcl-2 and pro-apoptotic protein Bim. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*, 1863(4), 650–659. <https://doi.org/10.1016/j.bbamcr.2016.01.006>
55. Menon, M. B., & Dhamija, S. (2018). Beclin 1 phosphorylation: At the center of autophagy regulation. *Frontiers in Cell and Developmental Biology*, 6, 137. <https://doi.org/10.3389/fcell.2018.00137>
56. Oral, O., Akkoc, Y., Bayraktar, O., & Gozuacik, D. (2016). Physiological and pathological significance of the molecular crosstalk between autophagy and apoptosis. *Histology and Histopathology*, 31(5), 479–498. <https://doi.org/10.14670/HH-11-714>
57. Yuan, Y., Zhang, Y., Han, L., Sun, S., & Shu, Y. (2018). miR-183 inhibits autophagy and apoptosis in gastric cancer cells by targeting ultraviolet radiation resistance-associated gene. *International Journal of Molecular Medicine*, 42(6), 3562–3570. <https://doi.org/10.3892/ijmm.2018.3871>
58. Murrow, L., Malhotra, R., & Debnath, J. (2015). ATG12-ATG3 interacts with Alix to promote basal autophagic flux and late endosome function. *Nature Cell Biology*, 17(3), 300–310. <https://doi.org/10.1038/ncb3112>
59. Tsai, T. C., Lai, K. H., Su, J. H., Wu, Y. J., & Sheu, J. H. (2018). 7-Acetylsinumaximol B induces apoptosis and autophagy in human gastric carcinoma cells through mitochondria dysfunction and activation of the PERK/eIF2 α /ATF4/CHOP signaling pathway. *Marine Drugs*, 16(4), 104. <https://doi.org/10.3390/md16040104>
60. Alway, S. E. (2019). Antioxidants and polyphenols mediate mitochondrial muscle death signaling in sarcopenia. In *Nutrition and Skeletal Muscle* (pp. 439–494). Academic Press. <https://doi.org/10.1016/B978-0-12-810422-4.00026-9>
61. Su, M., Mei, Y., & Sinha, S. (2013). Role of the crosstalk between autophagy and apoptosis in cancer. *Journal of Oncology*, 2013, 102735. <https://doi.org/10.1155/2013/102735>
62. Fairlie, W. D., Tran, S., & Lee, E. F. (2020). Crosstalk between apoptosis and autophagy signaling pathways. In *International Review of Cell and Molecular Biology* (Vol. 352, pp. 115–158). Elsevier. <https://doi.org/10.1016/bs.ircmb.2020.01.003>
63. McComb, S., Chan, P. K., Guinot, A., Hartmannsdottir, H., Jenni, S., Dobay, M. P., Bourquin, J. P., & Bornhauser, B. C. (2019). Efficient apoptosis requires feedback

- amplification of upstream apoptotic signals by effector caspase-3 or caspase-7. *Science Advances*, 5(7), eaau9433. <https://doi.org/10.1126/sciadv.aau9433>
64. Thermo Fisher Scientific. (n.d.). *Tracking autophagy with LC3B & p62*. Retrieved from <http://www.thermofisher.com/in/en/home/life-science/cell-analysis/cell-viability-and-regulation/autophagy/autophagy-lc3b-p62.html>
 65. Ojha, R., Ishaq, M., & Singh, S. (2015). Caspase-mediated crosstalk between autophagy and apoptosis: Mutual adjustment or matter of dominance. *Journal of Cancer Research and Therapeutics*, 11(3), 514–524. <https://doi.org/10.4103/0973-1482.163695>
 66. Tsapras, P., & Nezis, I. P. (2017). Caspase involvement in autophagy. *Cell Death & Differentiation*, 24(8), 1369–1379. <https://doi.org/10.1038/cdd.2017.43>
 67. Young, M. M., Takahashi, Y., Khan, O., Park, S., Hori, T., Yun, J., Sharma, A. K., Amin, S., Hu, C. D., Zhang, J., Kester, M., & Wang, H. G. (2012). Autophagosomal membrane serves as platform for intracellular death-inducing signaling complex (iDISC)-mediated caspase-8 activation and apoptosis. *Journal of Biological Chemistry*, 287(15), 12455–12468. <https://doi.org/10.1074/jbc.M111.309104>
 68. Liu, T., Zhang, J., Li, K., Deng, L., & Wang, H. (2020). Combination of an autophagy inducer and an autophagy inhibitor: A smarter strategy emerging in cancer therapy. *Frontiers in Pharmacology*, 11, 408. <https://doi.org/10.3389/fphar.2020.00408>
 69. Fitzwalter, B. E., Towers, C. G., Sullivan, K. D., Andrysiak, Z., Hoh, M., Ludwig, M., O'Prey, J., Ryan, K. M., Espinosa, J. M., Morgan, M. J., & Thorburn, A. (2018). Autophagy inhibition mediates apoptosis sensitization in cancer therapy by relieving FOXO3a turnover. *Developmental Cell*, 44(5), 555–565.e3. <https://doi.org/10.1016/j.devcel.2018.02.014>
 70. Chen, M. C., Lin, Y. C., Liao, Y. H., Liou, J. P., & Chen, C. H. (2019). MPT0G612, a novel HDAC6 inhibitor, induces apoptosis and suppresses IFN- γ -induced programmed death-ligand 1 in human colorectal carcinoma cells. *Cancers*, 11(10), 1617. <https://doi.org/10.3390/cancers11101617>
 71. Yu, H., Wu, C. L., Wang, X., Ban, Q., Quan, C., Liu, M., Dong, H., Li, J., Kim, G. Y., Choi, Y. H., Wang, Z., & Jin, C. Y. (2019). SP600125 enhances C-2-induced cell death by the switch from autophagy to apoptosis in bladder cancer cells. *Journal of Experimental & Clinical Cancer Research*, 38(1), 448. <https://doi.org/10.1186/s13046-019-1467-6>
 72. Tiliya Pun, N., Jang, W. J., & Jeong, C. H. (2020). Role of autophagy in regulation of cancer cell death/apoptosis during anti-cancer therapy: Focus on autophagy flux blockade. *Archives of Pharmacol Research*, 43(5), 475–488. <https://doi.org/10.1007/s12272-020-01239-w>
 73. Peng, Y., Qiu, L., Xu, D., Zhang, L., Yu, H., Ding, Y., Deng, L., & Lin, J. (2017). M4IDP, a zoledronic acid derivative, induces G1 arrest, apoptosis, and autophagy in HCT116 colon carcinoma cells via blocking the PI3K/Akt/mTOR pathway. *Life Sciences*, 185, 63–72. <https://doi.org/10.1016/j.lfs.2017.07.024>
 74. Kaluzki, I., Hailemariam-Jahn, T., Doll, M., Kaufmann, R., Balermipas, P., Zöller, N., Kippenberger, S., & Meissner, M. (2019). Dimethyl fumarate inhibits colorectal carcinoma cell proliferation: Evidence for cell cycle arrest, apoptosis, and autophagy. *Cells*, 8(11), 1329. <https://doi.org/10.3390/cells8111329>

75. Gugnoni, M., Sancisi, V., Manzotti, G., Gandolfi, G., & Ciarrocchi, A. (2016). Autophagy and epithelial–mesenchymal transition: An intricate interplay in cancer. *Cell Death & Disease*, 7(12), e2520. <https://doi.org/10.1038/cddis.2016.415>
76. Dang, S., Yu, Z., Zhang, C., Zheng, J., Li, K., Wu, Y., Qian, L., Yang, Z., Li, X., Zhang, Y., & Wang, R. (2015). Autophagy promotes apoptosis of mesenchymal stem cells under inflammatory microenvironment. *Stem Cell Research & Therapy*, 6, 247. <https://doi.org/10.1186/s13287-015-0245-4>
77. Lin, Y., Jiang, D., Li, Y., Han, X., Yu, D., Park, J. H., & Jin, Y. H. (2015). Effect of sun ginseng potentiation on epirubicin- and paclitaxel-induced apoptosis in human cervical cancer cells. *Journal of Ginseng Research*, 39(1), 22–28. <https://doi.org/10.1016/j.jgr.2014.08.001>
78. Hong, X., Li, S., Li, W., Xie, M., Wei, Z., Guo, H., Wei, W., & Zhang, S. (2019). Disruption of protein neddylation with MLN4924 attenuates paclitaxel-induced apoptosis and microtubule polymerization in ovarian cancer cells. *Biochemical and Biophysical Research Communications*, 508(4), 986–990. <https://doi.org/10.1016/j.bbrc.2018.12.048>
79. Deng, L., Wu, X., Zhu, X., Yu, Z., Liu, Z., Wang, J., & Zheng, Y. (2021). Combination effect of curcumin with docetaxel on the PI3K/AKT/mTOR pathway to induce autophagy and apoptosis in esophageal squamous cell carcinoma. *American Journal of Translational Research*, 13(1), 57–72.
80. Zhao, P., Li, M., Wang, Y., Chen, Y., He, C., Zhang, X., Yang, T., Lu, Y., You, J., Lee, R. J., & Xiang, G. (2018). Enhancing anti-tumor efficiency in hepatocellular carcinoma through autophagy inhibition by miR-375/sorafenib in lipid-coated calcium carbonate nanoparticles. *Acta Biomaterialia*, 72, 248–255. <https://doi.org/10.1016/j.actbio.2018.03.040>
81. Lin, Y., Jiang, D., Li, Y., Han, X., Yu, D., Park, J. H., & Jin, Y. H. (2015). Effect of sun ginseng potentiation on epirubicin- and paclitaxel-induced apoptosis in human cervical cancer cells. *Journal of Ginseng Research*, 39(1), 22–28. <https://doi.org/10.1016/j.jgr.2014.08.001>
82. Hong, X., Li, S., Li, W., Xie, M., Wei, Z., Guo, H., Wei, W., & Zhang, S. (2019). Disruption of protein neddylation with MLN4924 attenuates paclitaxel-induced apoptosis and microtubule polymerization in ovarian cancer cells. *Biochemical and Biophysical Research Communications*, 508(4), 986–990. <https://doi.org/10.1016/j.bbrc.2018.12.048>
83. Deng, L., Wu, X., Zhu, X., Yu, Z., Liu, Z., Wang, J., & Zheng, Y. (2021). Combination effect of curcumin with docetaxel on the PI3K/AKT/mTOR pathway to induce autophagy and apoptosis in esophageal squamous cell carcinoma. *American Journal of Translational Research*, 13(1), 57–72.
84. Zhao, P., Li, M., Wang, Y., Chen, Y., He, C., Zhang, X., Yang, T., Lu, Y., You, J., Lee, R. J., & Xiang, G. (2018). Enhancing anti-tumor efficiency in hepatocellular carcinoma through autophagy inhibition by miR-375/sorafenib in lipid-coated calcium carbonate nanoparticles. *Acta Biomaterialia*, 72, 248–255. <https://doi.org/10.1016/j.actbio.2018.03.040>